

Supplementary Table 1. Primers for the detection of bacterial groups

Eubacteria (all bacteria)¹

UniF340-Forward: 5'-ACTCCTACGGGAGGCAGCAGT-3'

UniR514-Reverse: 5'-ATTACCGCGGCTGCTGGC-3'

Ruminococcaceae (This study)

Forward: 5'-ATACCCTGGTAGTCCACGCT-3'

Reverse: 5'-GCGTCACTGGGATGTCAAGA-3'

Eubacterium rectale²

UniF338-Forward: 5'-ACTCCTACGGGAGGCAGC-3'

C.cocR491-Reverse: 5'-GCTTCTTAGTCAGGTACCGTCAT-3'

Lachnospiraceae (This study)

Forward: 5'-TGGATCCGCGTCTGATTAGC-3'

Reverse: 5'-TGTCTCAGTCCCAATGTGGC-3'

Lactobacillus group³

LabF362-Forward: 5'-AGCAGTAGGGAATCTTCCA-3'

LabR677-Reverse: 5'-CACCGCTACACATGGAG-3'

Segmented filamentous bacteria-1⁴

SFB736F-Forward: 5'-GACGCTGAGGCATAGCAT-3'

SFB844R-Reverse: 5'-GACGGCACGGATTGTTATTCA-3'

Segmented filamentous bacteria-2⁵

SFB1F 5'-AGGAGGAGTCTGCGGCAC-3'

SFB2R 5'-CCTTCCTCTCCCTGCT-3'

Clostridium coccooides⁶

Forward-5'-AAATGACGGTACCTGACTAA-3'

Reverse-5'-CTTTGAGTTTCATTCTTGCGAA-3'

Clostridium leptum (This study)

Forward: 5'-CCTTCCGTGCCGSAGTTA-3'

Reverse: 5'-GAATTA AACACATACTCCACTGCTT-3'

Turcibacter sp. (This study)

Forward: 5'-ACGGGGACAACGATTGGAAA-3'

Reverse: 5'-AGTGATGCCAGGAGCATCTT-3'

Bacteroides sp.⁷

BactF285-Forward: 5'-GGTTCTGAGAGGAGGTCCC-3'

UniR338-Reverse: 5'-GCTGCCTCCCGTAGGAGT-3'

Anaerophaga sp. (This study)

Forward: 5'-TCTCCGTGGGTTTAAAGGAAG-3'

Reverse: 5'-CGTGTCTCAGTACCAGTGTG-3'

Paludibacter sp. (This study)

Forward: 5'-AAGGAAGCGATTCCGGCTAC-3'

Reverse: 5'-CTCAGAACCCCTACGCATCG-3'

Prevotella sp.³

Forward- 5'-GGTGTCGGCTTAAGTGCCAT-3'

Reverse- 5'-CGGACGTAAGGGCCGTGC-3'

Helicobacter pylori⁸

Hp 547F-Forward: 5'-CTTAACCATAGAACTGCATTTGAAACTAC-3'

Hp 665R-Reverse: 5'-GGTCGCCTTCGCAATGAGTA-3'

Enterobacteriaceae⁹

Uni515F-Forward- 5'-GTGCCAGCAGCCGCGGTAA-3'

Ent826R-Reverse- 5'-GCCTCAAGGGCACAACCTCCAAG-3'

Akkermanisa muciniphila (This study)

Forward- 5'-CAGCACGTGAAGGTGGGGAC-3'

Reverse- 5'-CCTTGCGGTTGGCTTCAGAT-3'

The primer sets were either synthesized according to previous publications or designed based on DNA sequence information available in Ribosomal Database Project¹⁰ and validated.

**Supplementary Table 2. Pyrosequencing of 16S rRNA genes in the stool of diabetic
NOD mice**

Number of NOD mice tested	5
Number of sequences analyzed	5784 ± 1223
Number of phyla examined	4.2 ± 0.8
Number of genera investigated	21.8 ± 1.3
Number of OTUs evaluated	1300
Phylum percentages:	
<i>Firmicutes</i>	72.9 ± 11.9
<i>Bacteroidetes</i>	26.7 ± 11.9
<i>Tenericutes</i>	0.12 ± 0.2
<i>Actinobacteria</i>	0.08 ± 0.09
<i>Proteobacteria</i>	0.04 ± 0.03
Ratio of <i>Firmicutes/Bacteroidetes</i> :	3.4 ± 1.9
Class percentages:	
<i>Clostridia</i>	70.4 ± 10.93
<i>Bacteroidia</i>	26.8 ± 11.94
<i>Bacilli</i>	1.8 ± 0.86
<i>Erysipelotrichi</i>	0.42 ± 0.33
<i>Mollicutes</i>	0.12 ± 0.25
<i>Actinobacteria</i>	0.08 ± 0.09
<i>Alphaproteobacteria</i>	0.02 ± 0.02
<i>Gammaproteobacteria</i>	0.01 ± 0.03
Order percentages:	
<i>Clostridiales</i>	70.74 ± 4.8
<i>Bacteroidales</i>	26.76 ± 5.3
<i>Lactobacillales</i>	1.8 ± 0.38
<i>Erysipelotrichales</i>	0.42 ± 0.14
<i>Rhizobiales</i>	0.22 ± 0.01
<i>Anaeroplasmatales</i>	0.12 ± 0.11
<i>Micrococcales</i>	0.08 ± 0.04
<i>Enterobacteriales</i>	0.01 ± 0.01
<i>Rhodospirillales</i>	0.003 ± 0.003
Family percentages:	
<i>Lachnospiraceae</i>	64.35 ± 11.77

<i>Rikenellaceae</i>	18.86 ± 14.42
<i>Porphyromonadaceae</i>	9.99 ± 3.74
<i>Ruminococcaceae</i>	3.56 ± 1.05
<i>Lactobacillaceae</i>	1.79 ± 0.87
<i>Prevotellaceae</i>	0.56 ± 0.45
<i>Erysipelotrichaceae</i>	0.40 ± 0.33
<i>Anaeroplasmataceae</i>	0.16 ± 0.25
<i>Bacteroidaceae</i>	0.15 ± 0.09
<i>Micrococcaceae</i>	0.09 ± 0.09
<i>Clostridiaceae</i>	0.04 ± 0.07
<i>Methylobacteriaceae</i>	0.03 ± 0.02
<i>Enterobacteriaceae</i>	0.02 ± 0.04
<i>Rhodospirillaceae</i>	0.003 ± 0.003
Species percentages:	
<i>Firmicutes_Clostridia_Clostridiales_Lachnospiraceae_Lachnospiraceae</i> sp.	66.58 ± 11.7
<i>Firmicutes_Clostridia_Clostridiales_Lachnospiraceae_Coprococcus</i> sp.	0.33 ± 0.11
<i>Firmicutes_Clostridia_Clostridiales_Lachnospiraceae_Roseburia</i> sp.	0.31 ± 0.06
<i>Firmicutes_Clostridia_Clostridiales_Ruminococcaceae_Ruminococcaceae</i> sp.	2.01 ± 0.31
<i>Firmicutes_Clostridia_Clostridiales_Ruminococcaceae_Ruminococcus</i> sp.	0.22 ± 0.09
<i>Firmicutes_Clostridia_Clostridiales_Ruminococcaceae_Anaerotruncus</i> sp.	0.46 ± 0.11
<i>Firmicutes_Clostridia_Clostridiales_Ruminococcaceae_Faecalibacterium</i> sp.	0.01 ± 0.01
<i>Firmicutes_Bacilli_Lactobacillales_Lactobacillaceae_Lactobacillus johnsonii</i>	0.86 ± 0.33
<i>Firmicutes_Bacilli_Lactobacillales_Lactobacillaceae_Lactobacillus</i> sp.	0.55 ± 0.20
<i>Firmicutes_Bacilli_Lactobacillales_Lactobacillaceae_Lactobacillus reuteri</i>	0.39 ± 0.1
<i>Firmicutes_Clostridia_Clostridiales_Clostridiaceae_Clostridium_Clostridium</i> sp.	0.05 ± 0.03
<i>Firmicutes_Clostridia_Clostridiales_Oscillospiraceae_Oscillibacter</i> sp.	0.63 ± 0.05
<i>Firmicutes_Erysipelotrichia_Erysipelotrichales_Erysipelotrichaceae_Turcibacter</i> sp.	0.34 ± 0.15
<i>Bacteroidetes_Bacteroidia_Bacteroidales_Rikenellaceae_Alistipes</i> sp.	16.2 ± 6.4
<i>Bacteroidetes_Bacteroidia_Bacteroidales_Prevotellaceae_Prevotella</i> sp.	0.33 ± 0.17
<i>Bacteroidetes_Bacteroidia_Bacteroidales_Prevotellaceae_Prevotellaceae</i> sp.	0.14 ± 0.03
<i>Bacteroidetes_Bacteroidia_Bacteroidales_Prevotellaceae_Paraprevotella</i> sp.	0.04 ± 0.01
<i>Bacteroidetes_Bacteroidia_Bacteroidales_Bacteroidaceae_Bacteroides</i> sp.	0.13 ± 0.04
<i>Bacteroidetes_Bacteroidia_Bacteroidales_Barnesiellaceae_Barnesiella</i> sp.	8.86 ± 1.6
<i>Bacteroidetes_Bacteroidia_Bacteroidales_Odoribacteraceae_Odoribacter</i> sp.	1.19 ± 0.14
<i>Tenericutes_Mollicutes_Anaeroplasmatales_Anaeroplasmataceae_Anaeroplasma</i> sp.	0.12 ± 0.11
<i>Actinobacteria_Actinobacteria_Micrococcales_Micrococcaceae_Arthrobacter</i> sp.	0.08 ± 0.04
<i>Proteobacteria_Alphaproteobacteria_Rhizobiales_Methylobacteriaceae_Methylobacterium</i> sp.	0.02 ± 0.01
<i>Proteobacteria_Gammaproteobacteria_Enterobacterales_Enterobacteriaceae_Escherichia/Shigella</i> sp.	0.01 ± 0.01
unclassified sequences_metagenomes_organismal metagenomes_wallaby gut metagenome	0.01 ± 0.01

Phylum-Class-Order-Family-Genus-Species are listed according to the NCBI Taxonomy browser:

<https://www.ncbi.nlm.nih.gov/Taxonomy/Browser/wwwtax.cgi>. The DNA was derived from the stools of 28-30-wk old diabetic female NOD mice and subject to pyrosequencing individually using the bTEFAP method¹¹. The data were analyzed using a proprietary analysis pipeline by MR DNA Lab. Data shown are mean \pm SD of 5 samples.

Supplementary References

1. Amann RI, *et al.* Combination of 16S rRNA-targeted oligonucleotide probes with flow cytometry for analyzing mixed microbial populations. *Appl Environ Microbiol* 1990; 56:1919-25.
2. Franks AH, *et al.* Variations of bacterial populations in human feces measured by fluorescent in situ hybridization with group-specific 16S rRNA-targeted oligonucleotide probes. *Appl Environ Microbiol* **64**, 3336-3345 (1998).
3. Rinttilä T, *et al.* A. Development of an extensive set of 16S rDNA-targeted primers for quantification of pathogenic and indigenous bacteria in faecal samples by real-time PCR. *J Appl Microbiol* 2004; 97:1166-77.
4. Barman M, *et al.* Enteric salmonellosis disrupts the microbial ecology of the murine gastrointestinal tract. *Infect Immun* 2008; 76:907-15.
5. Dalby, AB, Frank DN, St Amand, AL. Culture-independent analysis of indomethacin-induced alterations in the rat gastrointestinal microbiota *Appl Environ Microbiol* 2006; 72, 6707-15.
6. Matsuki T, *et al.*, Use of 16S rRNA gene-targeted group-specific primers for real-time PCR analysis of predominant bacteria in human feces. *Appl Environ Microbiol* 2004; 70:7220-28.
7. Doré J, *et al.* Design and evaluation of a 16S rRNA-targeted oligonucleotide probe for specific detection and quantitation of human faecal Bacteroides populations. *Syst Appl Microbiol* 1998; 21:65-71.
8. Tan MP, *et al.* Chronic Helicobacter pylori infection does not significantly alter the microbiota of the murine stomach. *Appl Environ Microbiol* 2007; 73:1010-13.

9. Lane, D.J. rRNA sequencing: Nucleic acid techniques in bacterial systematics. In John Wiley and Sons, New York, pp. 115-175 (1991).
10. Cole JR, et al. Ribosomal Database Project: data and tools for high throughput rRNA analysis. *Nucleic Acids Res* 2014; 42:D633-42.
11. Dowd SE, et al. Bacterial tag-encoded FLX amplicon pyrosequencing (bTEFAP) for microbiome studies: bacterial diversity in the ileum of newly weaned Salmonella-infected pigs. *Foodborne Pathog Dis* 2008; 5:459-472.